

The host range of multi-host endophytic fungi

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Mature leaves of 224 angiosperm plant species belonging to 60 families and growing in Andaman Islands, and the states of Arunachal Pradesh, Kerala and Tamil Nadu were sampled for the presence of endophytic fungi. Fungal genera such as *Alternaria*, *Arthrinium*, *Aureobasidium*, *Chaetomium*, *Cladosporium*, *Glomerella/Colletotrichum*, *Drechslera*, *Fusarium*, *Fusicoccum*, *Lasiodiplodia*, *Paecilomyces*, *Pestalotiopsis*, *Phoma*, *Diaporthe/Phomopsis*, *Guignardia/Phyllosticta*, *Sporormiella* and *Xylaria* showed an isolation frequency of 5% or more. Species of *Colletotrichum*, *Phyllosticta*, *Phomopsis* and *Xylaria* occurred as endophytes in the leaves of many plant hosts including those that were taxonomically not closely related. The need to address the broad host range of some genera of fungal endophytes is discussed.

Keywords: Diversity, foliar endophytes, fungal endophytes, mutualism.

THE horizontally transmitted endophytic fungi, an integral part of the plant microbiome, infect the above and below ground tissues of all plants without eliciting any disease symptoms. A majority of those endophytes residing in the leaf belong to the subdivision Ascomycotina and are globally distributed in plants growing from the Arctic¹ to the Antarctica². The endophytic lifestyle is exhibited by various ecological groups of fungi including mutualists, latent pathogens and saprotrophs³⁻⁵. Although universal in occurrence, information regarding endophyte biology such as evolution of endophytism as a lifestyle among fungi, interactions of endophytes with their host plants and other organisms associated with the host plants as well as with the environment is inadequate⁶. For

instance, though many independent studies show that certain genera including *Colletotrichum*, *Guignardia* (*Phyllosticta*), *Pestalotiopsis*, *Diaporthe* (*Phomopsis*), and *Xylaria* occur as endophytes in a wide range of plants from different geographical locations, principally, their host range has not been addressed.

Here, using the data gathered in our 20-year long-term study on foliar endophytes of plants from various parts of India, we analyse the distribution of some of the more commonly encountered endophyte genera. The study includes 224 angiosperm plant species belonging to 60 families and were collected from Andaman Islands (Shoal bay – 11°53'52.53"N, 92°46'32.1204"E, Wright myo – 11°47'19.0896"N, 92°43'34.3884"E, Shippighat – 11°39'52.3368"N, 92°44'11.04"E, Corbyn's cove – 11°38'40.7976"N, 92°44'51.3636"E, Manjeri – 11°32'33.36"N, 92°39'8.7588"E, Burmanallah – 11°33'27.3924"N, 92°43'47.3448"E and Chidiyatapu – 11°30'21.9564"N, 92°42'6.0948"E), and the states of Arunachal Pradesh (Ronohills – 27°9'9.702"N, 93°45'56.8116"E), Kerala (Peechi – 10°31'37.3368"N, 76°21'39.0024"E) and Tamil Nadu (Chennai – 13°4'57.648"N, 80°16'14.5848"E, Chengalpattu – 12°41'2.382"N, 79°59'0.0528"E, Pichavaram – 11°25'54.7752"N, 79°46'51.7692"E, Nilgiri Biosphere Reserve – Masinagudi – 11°34'19.8372"N, 76°38'33.774"E, Kargudi and Upper Kargudi – 11°34'30.036"N, 76°33'19.5768"E and Upper Bhavani – 11°13'47.1504"N, 76°31'24.7512"E) and included a wide range and variety of hosts including mangroves, hydrophytes and mesophytes.

The methodologies used, such as the criteria for sampling, surface sterilization procedure, size of plant tissue segment screened, growth medium and incubation procedure for isolation of endophytes were the same for all the plants studied⁷ so that the data obtained could be compared. To present the results, we consider only those endophyte isolates that appeared in 5% or more of the tissue bits plated for each plant species.

We observed that 17 genera of fungi, viz. *Alternaria*, *Arthrinium*, *Aureobasidium*, *Chaetomium*, *Cladosporium*, *Glomerella/Colletotrichum*, *Drechslera*, *Fusarium*, *Fusicoccum*, *Lasiodiplodia*, *Paecilomyces*, *Pestalotiopsis*, *Phoma*, *Diaporthe/Phomopsis*, *Guignardia/Phyllosticta*, *Sporormiella* and *Xylaria* exhibited an isolation frequency of 5% or more in many plant hosts (Table 1). A comparison of the data showed that genera such as *Colletotrichum*, *Phyllosticta*, *Phomopsis* and *Xylaria* occur as endophytes in the leaves of plants of disparate families. Of the 224 plant species screened, 167 harboured *Phomopsis* and *Phyllosticta* spp. as endophytes whereas species of *Colletotrichum* and *Xylaria* were present in the leaves of 160 and 124 plant species respectively (Table 1). These four genera were the most common foliar endophytes and could be isolated from 55% to 75% of the plant species screened (Table 1). *Colletotrichum* was endophytic in both members of Lamiaceae and

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Table 1. Endophyte genera (isolation % >5 at least in one of the plant species studied) isolated from leaves of different angiosperm hosts

Family	Host	Location	Dominant endophytes																
			a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	p	q
Acanthaceae (7)	<i>Acanthus ebracteatus</i>	Shippighat	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Acanthus ilicifolius</i>	Wright myo	○	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Andrographis echinoides</i>	Chengalpattu	○	○	○	•	○	○	○	•	•	○	○	○	○	○	○	○	○
	<i>Avicennia marina</i>	Pichavaram	○	•	○	•	○	•	○	○	○	○	•	○	○	○	○	○	○
	<i>Avicennia officinalis</i>	Pichavaram	○	○	○	○	○	•	○	○	○	•	○	○	○	○	○	○	○
	<i>Justicia adhatoda</i>	Chengalpattu	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Justicia gendarussa</i>	Chengalpattu	○	○	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○
Altingiaceae (1)	<i>Altingia excelsa</i>	Ronohills	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	
Anacardiaceae (3)	<i>Anacardium occidentale</i>	Chennai	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Mangifera indica</i>	Chennai	•	•	•	•	○	○	○	○	○	○	○	•	•	○	○	○	
	<i>Semecarpus anacardium</i>	Masinagudi	•	•	•	•	•	•	○	•	•	○	○	○	○	○	○	○	
Apocynaceae (16)	<i>Allamanda cathartica</i>	Chennai	○	○	○	•	○	•	○	○	○	○	•	○	○	○	○	○	
	<i>Calotropis gigantea</i>	Chennai	•	○	○	•	○	○	•	○	○	○	○	○	○	•	○	○	
	<i>Cascabela thevetia</i>	Chennai	○	•	○	•	○	•	○	○	○	○	○	○	○	○	○	○	
	<i>Catharanthus roseus</i>	Chennai	○	•	○	•	○	○	○	○	○	○	○	○	○	•	○	○	
	<i>Cryptostegia grandiflora</i>	Chennai	○	•	○	•	○	•	○	○	○	○	○	○	○	○	○	○	
	<i>Gymnema sylvestre</i>	Chennai	○	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Hemidesmus indicus</i>	Chennai	○	•	○	•	○	○	○	○	○	○	•	○	○	•	○	○	
	<i>Nerium oleander</i>	Chennai	•	•	○	○	○	•	○	○	○	○	○	○	○	○	○	○	
	<i>Oxystelma secamone</i>	Chennai	○	•	○	•	○	•	○	○	○	○	○	○	•	•	○	○	
	<i>Pentatropis capensis</i>	Chennai	•	•	○	•	○	○	•	○	○	○	○	○	○	•	○	○	
	<i>Plumeria rubra</i>	Chennai	•	•	○	•	•	•	○	○	○	○	○	○	○	○	○	○	
	<i>Rauvolfia tetraphylla</i>	Chennai	○	•	○	○	○	○	○	○	○	○	○	○	○	•	○	○	
	<i>Sarcostemma acidum</i>	Chennai	•	○	○	•	○	○	•	•	○	○	•	○	○	•	○	○	
	<i>Tabernaemontana divaricata</i>	Chennai	•	•	○	•	○	○	•	○	○	○	○	○	○	○	○	○	
	<i>Tylophora indica</i>	Chennai	•	•	○	•	○	○	○	○	○	○	○	○	○	•	○	○	
	<i>Wattakaka volubilis</i>	Chennai	○	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	
Aquifoliaceae (2)	<i>Ilex denticulata</i>	Upper Bhavani	•	○	○	•	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Ilex wightiana</i>	Upper Bhavani	○	○	○	○	•	•	○	○	○	○	○	○	○	○	○	○	
Araceae (1)	<i>Pistia stratiotes</i>	Chennai	○	○	○	○	○	•	○	○	•	○	○	○	○	○	○	•	
Araliaceae (1)	<i>Heteropanax fragrans</i>	Ronohills	○	○	○	•	○	○	○	○	○	○	○	○	○	○	○	○	
Arecaceae (32)	<i>Areca catechu</i>	Peechi	○	○	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Borassus flabellifer</i>	Chennai	○	○	•	○	○	○	•	○	•	○	○	○	○	○	○	○	
	<i>Calamus andamanicus</i>	Peechi	•	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus dransfieldii</i>	Peechi	○	•	○	○	○	○	○	○	○	○	•	○	○	○	○	○	
	<i>Calamus hookerianus</i>	Peechi	•	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus metzianus</i>	Peechi	○	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus nagbettai</i>	Peechi	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus pseudotenuis</i>	Peechi	•	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus rotang</i>	Peechi	•	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus tenuis</i>	Peechi	○	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus tetradactylus</i>	Peechi	•	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus thwaitesii</i>	Peechi	•	•	○	○	•	○	○	•	○	○	○	○	○	○	○	○	
	<i>Calamus travancoricus</i>	Peechi	○	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Calamus vattayila</i>	Peechi	•	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Caryota urens</i>	Chennai	•	•	○	•	•	○	•	○	○	○	○	○	○	○	○	○	
	<i>Chrysalidocarpus lutescens</i>	Chennai	○	○	○	○	○	•	○	○	○	○	○	○	•	○	○	○	
	<i>Cocos nucifera</i>	Chennai	•	•	○	○	○	○	•	○	○	○	○	○	•	○	○	○	
	<i>Corypha macropoda</i>	Chennai	•	○	○	○	•	○	○	○	○	○	○	•	○	○	○	○	
	<i>Cyrtostachys lakka</i>	Peechi	•	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Elaeis guineensis</i>	Chennai	•	•	○	○	○	•	•	○	•	○	○	○	•	○	○	○	
	<i>Licuala grandis</i>	Peechi	○	•	○	○	○	•	○	○	○	○	○	○	○	○	○	○	
	<i>Livistona chinensis</i>	Peechi	•	•	○	○	○	•	○	○	○	○	○	○	○	○	○	○	
	<i>Livistona jenkinsiana</i>	Ronohills	•	○	○	•	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Livistona rotundifolia</i>	Peechi	○	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Nypa fruticans</i>	Shippighat	•	○	•	•	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Phoenix paludosa</i>	Shippighat	•	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Phoenix rupicola</i>	Ronohills	○	○	○	•	○	○	○	○	•	○	○	○	○	○	○	○	
	<i>Phoenix sylvestris</i>	Chennai	○	•	○	•	○	○	•	○	○	○	○	○	○	○	○	○	
	<i>Pritchardia pacifica</i>	Chennai	○	○	○	○	•	•	•	○	○	○	•	○	○	○	○	○	

(Contd)

Table 1. (Contd)

Family	Host	Location	Dominant endophytes																
			a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	p	q
	<i>Ptychosperma macarthurii</i>	Chennai	○	●	○	○	○	○	●	○	○	○	○	○	○	○	○	○	○
	<i>Rhapis excelsa</i>	Chennai	○	○	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Roystonea regia</i>	Chennai	○	○	○	○	○	●	●	○	○	○	○	○	●	○	○	○	○
Asteraceae (1)	<i>Spilanthes oleracea</i>	Ronohills	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Bignoniaceae (4)	<i>Radermachera xylocarpa</i>	Kargudi	●	●	○	○	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Stereospermum angustifolium</i>	Masinagudi	●	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Stereospermum personatum</i>	Kargudi	●	●	●	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Stereospermum tetragonum</i>	Upper Kargudi	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Boraginaceae (4)	<i>Coldenia procumbens</i>	Chengalpattu	○	○	○	○	○	○	○	●	○	○	○	○	○	●	○	○	○
	<i>Cordia dichotoma</i>	Kargudi	●	●	○	○	○	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Cordia obliqua</i>	Masinagudi	●	●	○	●	●	○	○	○	●	○	○	○	○	○	○	○	○
	<i>Ehretia canarensis</i>	Masinagudi	●	●	○	●	●	○	○	●	●	○	○	○	○	○	○	○	○
Calophyllaceae (1)	<i>Mesua assamica</i>	Ronohills	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Caprifoliaceae (1)	<i>Viburnum punctatum</i>	Upper Kargudi	●	●	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○
Celastraceae (3)	<i>Elaeodendron glaucum</i>	Masinagudi	●	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Euonymus angulatus</i>	Upper Bhavani	●	○	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Maytenus emarginata</i>	Masinagudi	●	●	●	●	●	●	○	○	○	○	○	○	○	○	○	○	○
Colchicaceae (1)	<i>Gloriosa superba</i>	Chengalpattu	●	●	●	●	●	○	○	○	○	○	○	○	○	●	○	○	○
Combretaceae (8)	<i>Anogeissus latifolia</i>	Masinagudi	●	●	○	●	●	○	○	●	○	○	○	○	○	○	○	○	○
	<i>Lumnitzera littorea</i>	Shoal Bay	●	●	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Lumnitzera racemosa</i>	Manjeri	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Terminalia alata</i>	Kargudi	●	●	●	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Terminalia arjuna</i>	Ronohills	●	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Terminalia catappa</i>	Ronohills	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Terminalia chebula</i>	Masinagudi	●	●	●	○	●	●	○	○	●	○	○	○	○	○	○	○	○
	<i>Terminalia crenulata</i>	Kargudi	●	●	●	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Convolvulaceae (1)	<i>Ipomoea aquatica</i>	Chennai	●	○	○	●	○	○	○	●	○	○	○	○	○	○	○	○	○
Dioscoreaceae (1)	<i>Dioscorea</i> sp.	Ronohills	●	○	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Dipterocarpaceae (1)	<i>Shorea roxburghii</i>	Kargudi	●	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○
Ebenaceae (1)	<i>Diospyros montana</i>	Masinagudi	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Elaeocarpaceae (2)	<i>Elaeocarpus sphaericus</i>	Ronohills	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Elaeocarpus serratus</i>	Upper Bhavani	●	○	○	●	●	○	○	○	●	○	○	○	○	○	○	○	○
Elatinaceae (1)	<i>Bergia aquatica</i>	Chennai	○	○	○	●	○	○	○	●	○	○	○	○	○	○	○	○	○
Erythroxylaceae (1)	<i>Erythroxylum monogynum</i>	Masinagudi	●	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
Euphorbiaceae (10)	<i>Acalypha indica</i>	Chennai	●	●	○	●	○	○	○	○	○	○	○	○	○	●	○	○	○
	<i>Bridelia retusa</i>	Masinagudi	●	●	●	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Daphniphyllum neilgherrense</i>	Upper Bhavani	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Excoecaria agallocha</i>	Pichavaram	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Givotia rotleriformis</i>	Masinagudi	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Glochidion zeylanicum</i>	Upper Bhavani	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Macaranga peltata</i>	Upper Bhavani	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Mallostus paniculatus</i>	Ronohills	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Phyllanthus amarus</i>	Chennai	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Phyllanthus emblica</i>	Masinagudi	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Fabaceae (18)	<i>Acacia chundra</i>	Masinagudi	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Acacia ferruginea</i>	Masinagudi	●	●	○	○	○	●	○	●	●	○	○	○	○	○	○	○	○
	<i>Acacia leucophloea</i>	Masinagudi	●	●	○	○	○	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Acacia suma</i>	Masinagudi	●	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Aeschynomene aspera</i>	Chennai	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Albizia amara</i>	Masinagudi	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Albizia saman</i>	Ronohills	○	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Bauhinia racemosa</i>	Masinagudi	●	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Bauhinia variegata</i>	Chennai	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Butea monosperma</i>	Masinagudi	●	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Caesalpinia pulcherrima</i>	Chennai	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Cassia fistula</i>	Masinagudi	●	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Dalbergia lanceolaria</i>	Masinagudi	●	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Dalbergia latifolia</i>	Upper Kargudi	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○

(Contd)

RESEARCH COMMUNICATIONS

Table 1. (Contd)

Family	Host	Location	Dominant endophytes																	
			a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	p	q	
	<i>Ougeinia oojeinensis</i>	Kargudi	•	•	•	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Pongamia pinnata</i>	Masinagudi	•	•	•	•	•	○	○	•	○	○	○	○	○	○	○	○	○	
	<i>Pterocarpus marsupium</i>	Masinagudi	•	•	○	•	•	•	○	•	•	○	○	○	○	○	○	○	○	
	<i>Samanea saman</i>	Masinagudi	•	•	○	○	•	•	○	○	○	○	○	○	○	○	○	○	○	
Staphyleaceae (1)	<i>Turpinia cochinchinensis</i>	Upper Bhavani	•	•	•	•	•	•	○	○	○	○	○	○	○	○	○	○		
Lamiaceae (10)	<i>Callicarpa arborea</i>	Ronohills	•	○	○	•	○	○	○	•	○	○	○	○	○	○	○	○	○	
	<i>Clerodendrum colebrookianum</i>	Ronohills	•	○	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Clerodendrum sp.</i>	Ronohills	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Gmelina arborea</i>	Kargudi	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Gmelina asiatica</i>	Masinagudi	•	•	•	•	•	○	•	○	•	○	○	○	○	○	○	○	○	
	<i>Leucas aspera</i>	Chengalpattu	○	○	○	○	○	○	○	○	○	○	•	○	○	○	•	○	○	○
	<i>Premna tomentosa</i>	Masinagudi	•	•	○	•	•	•	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Tectona grandis</i>	Kargudi	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Vitex altissima</i>	Kargudi	•	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Vitex negundo</i>	Chengalpattu	•	•	○	•	○	○	○	○	•	○	○	○	○	○	•	○	○	○
Lauraceae (10)	<i>Cinnamomum malabratrum</i>	Upper Bhavani	•	•	•	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Cinnamomum wightii</i>	Upper Bhavani	•	○	○	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Cryptocarya bourdillonii</i>	Upper Bhavani	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Litsea cubeba</i>	Ronohills	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Litsea floribunda</i>	Upper Bhavani	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Litsea oleoides</i>	Upper Bhavani	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Litsea stocksii</i>	Upper Bhavani	•	○	○	•	•	○	○	○	•	○	○	○	○	○	○	○	○	○
	<i>Neolitsea zeylanica</i>	Upper Bhavani	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Persea macrantha</i>	Upper Kargudi	•	•	○	○	○	○	○	○	•	○	○	○	○	○	○	○	○	○
	<i>Phoebe lanceolata</i>	Upper Bhavani	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	○
Lecythidaceae (1)	<i>Careya arborea</i>	Kargudi	•	•	•	○	○	○	○	○	•	○	○	○	○	○	○	○	○	
Loganiaceae (1)	<i>Strychnos potatorum</i>	Masinagudi	•	•	○	•	•	•	○	•	•	○	○	○	○	○	○	○	○	
Lythraceae (4)	<i>Lagerstroemia microcarpa</i>	Kargudi	•	•	○	•	•	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Lagerstroemia parviflora</i>	Kargudi	•	•	•	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Lagerstroemia sp.</i>	Ronohills	○	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Sonneratia alba</i>	Wright myo	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
Magnoliaceae (1)	<i>Michelia nilagirica</i>	Upper Bhavani	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
Malvaceae (4)	<i>Grewia tiliifolia</i>	Kargudi	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Helicteres isora</i>	Kargudi	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Kydia calycina</i>	Kargudi	•	•	○	•	○	○	○	○	•	○	○	○	○	○	○	○	○	
	<i>Sida acuta</i>	Ronohills	○	○	○	•	○	○	○	•	•	○	○	○	○	○	○	○	○	
Marantaceae (1)	<i>Phrynium capitatum</i>	Ronohills	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
Melastomataceae (1)	<i>Memecylon malabaricum</i>	Upper Bhavani	•	○	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
Meliaceae (1)	<i>Xylocarpus granatum</i>	Burmanallah	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
Meliosmaceae (1)	<i>Meliosma simplicifolia</i>	Upper Bhavani	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
Moraceae (2)	<i>Ficus benghalensis</i>	Chennai	•	•	○	○	•	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Ficus religiosa</i>	Ronohills	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
Myrsinaceae (1)	<i>Aegiceras corniculatum</i>	Corbyn's Cove	•	•	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
Myrtaceae (4)	<i>Callistemon viminalis</i>	Ronohills	•	○	○	•	○	○	○	•	○	○	○	○	○	○	○	○	○	
	<i>Rhodomyrtus tomentosa</i>	Upper Bhavani	•	•	○	•	•	○	○	○	•	○	○	○	○	○	○	○	○	
	<i>Syzygium cumini</i>	Kargudi	•	•	•	•	•	•	○	○	○	○	○	○	○	○	○	○	○	
	<i>Syzygium densiflorum</i>	Upper Bhavani	•	○	○	•	•	○	○	○	•	○	○	○	○	○	○	○	○	○
Nymphaeaceae (1)	<i>Nymphaea stellata</i>	Chennai	○	○	○	•	○	○	○	○	○	○	○	○	•	○	○	○	○	
Oleaceae (4)	<i>Ligustrum roxburghii</i>	Upper Bhavani	•	•	•	•	•	○	○	○	•	○	○	○	○	○	○	○	○	
	<i>Olea glandulifera</i>	Upper Kargudi	•	○	○	•	○	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Olea dioica</i>	Upper Kargudi	○	○	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Schrebera swietenoides</i>	Kargudi	•	•	○	•	○	•	○	○	•	○	○	○	○	○	○	○	○	
Orchidaceae (11)	<i>Aerides odorata</i>	Ronohills	•	•	○	•	•	○	○	○	○	○	○	○	•	○	○	○	○	
	<i>Arundina graminifolia</i>	Ronohills	○	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Cymbidium aloifolium</i>	Ronohills	○	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	
	<i>Cymbidium ensifolium</i>	Ronohills	•	•	○	•	•	○	○	○	○	○	○	○	○	○	○	○	○	

(Contd)

Table 1. (Contd)

Family	Host	Location	Dominant endophytes																
			a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	p	q
	<i>Dendrobium fimbriatum</i>	Ronohills	○	●	○	●	●	○	○	●	○	○	○	○	○	○	○	○	○
	<i>Dendrobium moschatum</i>	Ronohills	○	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Eria flava</i>	Ronohills	●	●	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Paphiopedilum fairrieianum</i>	Ronohills	○	●	○	○	●	○	○	○	○	○	○	○	●	○	○	○	○
	<i>Pholidota imbricata</i>	Ronohills	○	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Rhynchostylis retusa</i>	Ronohills	○	○	○	○	●	○	○	○	○	○	○	○	●	○	○	○	○
	<i>Vanilla planifolia</i>	Ronohills	○	●	○	○	●	○	○	○	○	○	○	○	●	○	○	○	○
Pandanaceae (1)	<i>Pandanus nepalensis</i>	Ronohills	●	●	●	●	○	○	○	○	●	○	○	○	○	○	○	○	○
Piperaceae (3)	<i>Piper betle</i>	Chennai	○	○	○	●	○	○	○	●	●	○	○	○	○	○	○	○	○
	<i>Piper mullesua</i>	Ronohills	●	●	○	●	○	○	○	●	○	○	○	○	○	○	○	○	○
	<i>Piper</i> sp.	Ronohills	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Pontederiaceae (2)	<i>Eichhornia crassipes</i>	Chennai	●	●	○	●	○	○	●	○	○	○	○	○	○	○	○	●	○
	<i>Monochoria hastifolia</i>	Chennai	●	○	○	●	○	○	○	○	●	○	○	○	○	●	○	●	○
Rhamnaceae (2)	<i>Ziziphus jujuba</i>	Masinagudi	●	●	○	●	●	○	○	○	●	○	○	○	○	○	○	○	○
	<i>Ziziphus xylopyrus</i>	Masinagudi	●	●	○	●	○	●	○	○	○	○	○	○	○	○	○	○	○
Rhizophoraceae (7)	<i>Bruguiera cylindrica</i>	Chidiyatapu	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Bruguiera gymnorrhiza</i>	Burmanallah	●	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Bruguiera parviflora</i>	Shoal Bay	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Ceriops tagal</i>	Manjeri	●	●	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Rhizophora apiculata</i>	Burmanallah	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Rhizophora mucronata</i>	Burmanallah	●	○	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Rhizophora stylosa</i>	Chidiyatapu	●	○	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Rubiaceae (8)	<i>Canthium parviflorum</i>	Masinagudi	●	●	○	●	●	●	○	●	○	○	○	○	○	○	○	○	○
	<i>Catunaregam spinosa</i>	Kargudi	○	○	○	●	○	○	○	●	●	○	○	○	○	○	○	○	○
	<i>Ixora coccinea</i>	Chennai	●	●	○	○	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Ixora nigricans</i>	Masinagudi	●	●	○	○	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Lasianthus venulosus</i>	Upper Bhavani	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Psychotria bisulcata</i>	Upper Bhavani	●	○	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Randia dumetorum</i>	Masinagudi	●	●	●	●	●	○	○	○	●	○	○	○	○	○	○	○	○
	<i>Scyphiphora hydrophylacea</i>	Shoal Bay	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Rutaceae (3)	<i>Murraya paniculata</i>	Upper Kargudi	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Naringi crenulata</i>	Masinagudi	●	●	○	●	●	●	○	○	○	○	○	○	○	○	○	○	○
	<i>Vepris bilocularis</i>	Upper Bhavani	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Salicaceae (2)	<i>Casearia esculenta</i>	Kargudi	●	●	○	●	○	○	○	●	○	○	○	○	○	○	○	○	○
	<i>Salix tetrasperma</i>	Upper Kargudi	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Sapindaceae (1)	<i>Sapindus emarginatus</i>	Masinagudi	●	●	●	●	●	●	○	●	●	○	○	○	○	○	○	○	○
Sapotaceae (1)	<i>Isonandra candolleana</i>	Upper Bhavani	●	●	○	●	○	●	○	○	○	○	○	○	○	○	○	○	○
Saururaceae (1)	<i>Houttuynia</i> sp.	Ronohills	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Scrophulariaceae (1)	<i>Scoparia dulcis</i>	Ronohills	●	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Solanaceae (5)	<i>Capsicum annum</i>	Chennai	●	●	○	●	○	○	●	○	○	○	○	○	○	○	○	○	○
	<i>Datura metel</i>	Chennai	○	●	○	●	○	○	○	●	○	○	○	○	○	●	○	○	○
	<i>Solanum melongena</i>	Chennai	○	●	○	●	○	○	○	○	○	○	○	○	●	●	○	○	○
	<i>Solanum torvum</i>	Ronohills	●	○	○	●	○	○	○	●	○	○	○	○	○	○	○	○	○
	<i>Withania somnifera</i>	Upper Kargudi	●	●	○	●	○	○	○	○	○	○	○	○	○	○	○	○	○
Symplocaceae (2)	<i>Symplocos cochinchinensis</i>	Upper Bhavani	●	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Symplocos obtusa</i>	Upper Bhavani	●	○	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○
Theaceae (2)	<i>Eurya nitida</i>	Upper Bhavani	○	●	○	○	●	○	○	○	○	○	○	○	○	○	○	○	○
	<i>Ternstroemia japonica</i>	Upper Bhavani	●	○	○	●	●	○	○	○	○	○	○	○	○	○	○	○	○
Ulmaceae (1)	<i>Holoptelea integrifolia</i>	Masinagudi	●	●	○	●	●	●	●	○	○	○	○	○	○	○	○	○	○
Urticaceae (1)	<i>Pouzolzia</i> sp.	Ronohills	○	●	○	●	●	○	○	○	○	○	○	○	○	○	○	○	●
Dominant endophyte genus			a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	p	q
No. of plant hosts harbouring the dominant endophyte genus			167	167	30	160	124	50	19	30	33	2	6	1	11	18	1	4	1
% of plant hosts harbouring the dominant endophyte genus			75	75	13	71	55	22	9	13	15	1	3	0.4	5	8	0.4	2	0.4

Number after family name indicates the number of species screened.

(a) *Phomopsis*; (b) *Phyllosticta*; (c) *Pestalotiopsis*; (d) *Colletotrichum*; (e) *Xylaria*; (f) *Sporormiella*; (g) *Aureobasidium*; (h) *Fusarium*; (i) *Lasiodiplodia*; (j) *Paecilomyces*; (k) *Phoma*; (l) *Fusicoccum*; (m) *Cladosporium*; (n) *Alternaria*; (o) *Chaetomium*; (p) *Dreschlera*; (q) *Arthrinium*. ●, Present, ○, Absent or <5% colonization frequency.

Nymphaeaceae, the most distantly related families screened as exemplified by the phylogenetic tree (Figure 1). Similarly, *Phomopsis* could be isolated from disparate families such as Scrophulariaceae and Pandanaeae (Figure 1).

The multi-host nature of some endophyte genera such as *Colletotrichum*, *Pestalotiopsis*, *Phomopsis*, *Phyllosticta* and *Xylaria* is well known^{5,8-13}. In the dry tropical forests, these endophytes are not only widespread but also dominate the endophyte assemblage of their plant hosts such that the overall endophyte diversity in these ecosystems is depressed⁷. The present study involving a large sample size attests the wide host range of such endophytes. Although one of the factors critical for a wide host range among pathogens is the high levels of genetic diversity among them¹⁴, with reference to endophytes, it is not known if the same species infecting different plants has evolved traits to overcome the different

defence mechanisms of such plant species. One of the *Pestalotiopsis* species recorded in the current study has been shown to escape detection by the plant immune system by modifying its cell wall chitin structure while infecting the plant¹⁵. In plant–fungus interaction, host sharing among fungi declines as a function of phylogenetic distance between plant hosts¹⁶ and thus closely related plants harbour the same fungal species. This is observed among conifer needle endophytes¹⁷ and tropical leaf pathogens¹⁸ but not for at least some foliar endophytic fungi owing to their multi-host nature. Such a wide host range reflects the high ecological amplitude of these endophytes, since a host plant’s defence compounds¹⁹ and nutrient content²⁰ determines the endophyte community supported by it. Furthermore, their competitive ability should also be high as they are bound to encounter constituents of different microbiomes associated with different plant hosts²¹.

It has been argued that generally, evolution should favour host restriction among pathogens. This is because the concomitant functional trade-offs could restrict the fitness of generalists in a given habitat and the possibility of slower evolution in the broad niche occupied by the generalists¹⁴. However, according to May^{22,23}, host specificity is low among plant-associated organisms in the tropics due to non-contiguous distribution of a plant host owing to the high plant diversity existing here. This possibly leads to the development of host generalism. Another possibility is the development of common traits by endophytic fungi to survive in unique environments leading to their selection and subsequent survival in different plant hosts. For instance, different endophyte species of dry tropical forests experiencing prolonged dry periods and episodic forest fires have common traits such as thermo-tolerant spores, tolerance to toxic furaldehydes which are released during biomass burning, and a biphasic life style (endophytic in intact leaves and saprobic in fallen litter)^{5,24,25}. Thus, host range expansion in these endophytes could be, apart from co-speciation, due to frequent host jumps as exemplified by some fungal endophytes and pathogens^{26,27}. The host could also play a significant role in determining the host range of endophytes. Redman *et al.*²⁸ showed that for some multi-host endophytes exhibiting both pathogenic and endophytic lifestyles, it is the plant host’s metabolism which determines if the invader should survive as a mutualist or a pathogen.

Endophytism is a successful lifestyle among fungi as it evolved about 400 million years ago²⁹ and is exhibited by several functional groups of fungi. Infection by multi-host and dominant leaf endophyte alters the expression of many defence-linked genes of the plant host³⁰. It is not known if prior infection by such an endophyte determines subsequent endophyte colonizers due to its effect on the plant’s defence status. Similarly, since there are trade-offs for the host plants even in the beneficial mycorrhizal association³¹, it would be worthwhile unravelling the cost

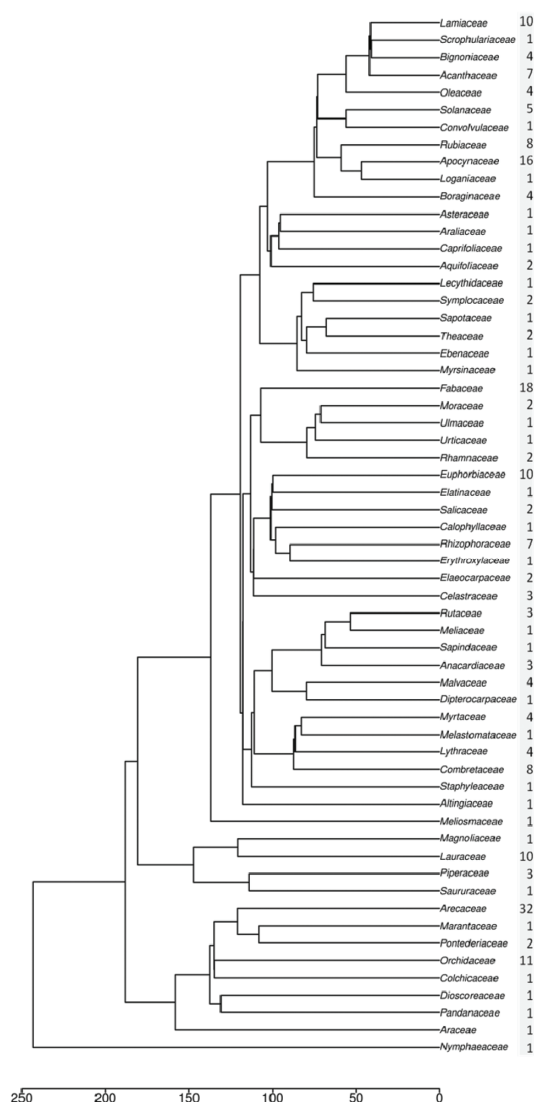


Figure 1. Phylogram of 60 angiosperm families screened for endophytes obtained by using the function S. PhyloMaker. Numbers indicate the number of plant species studied for each family.

borne by plants for harbouring such multi-host endophytes. Furthermore, since pathogens could survive as endophytes (latent pathogens)³, and since many species of *Phomopsis*³² and *Colletotrichum*³³ have a wide host range as symptomless endophytes, it is even more essential to explore the host acquiring ability of such multi-host endophytes. Collectively, to appreciate more completely the evolution of endophytism, it is essential to focus on the question as to how some fungal genera have evolved to become generalist endophytes at a global scale.

- Zhang, T. and Yao, Y. F., Endophytic fungal communities associated with vascular plants in the high arctic zone are highly diverse and host plant specific. *PLoS ONE*, 2015, **10**, e0130051.
- Rosa, L. H., Vaz, A. B. M., Caligiorno, R. B., Campolina, S. and Rosa, C. A., Endophytic fungi associated with the Antarctic grass *Deschampsia Antarctica* Desv. (Poaceae). *Polar Biol.*, 2009, **32**, 161–167.
- Suryanarayanan, T. S. and Murali, T. S., Incidence of *Leptosphaeria crassiasca* in symptomless leaves of peanut in southern India. *J. Basic Microbiol.*, 2006, **46**, 305–309.
- Davis, E. C. and Shaw, A. J., Biogeographic and phylogenetic patterns in diversity of liverwort-associated endophytes. *Am. J. Bot.*, 2008, **95**, 914–924.
- Sudhakara Reddy, M., Murali, T. S., Suryanarayanan, T. S., Govinda Rajulu, M. B. and Thirunavukkarasu, N., *Pestalotiopsis* species occur as generalist endophytes in trees of Western Ghats forests of southern India. *Fungal Ecol.*, 2016, **24**, 70–75.
- Suryanarayanan, T. S., Endophyte research: going beyond isolation and metabolite documentation. *Fungal Ecol.*, 2013, **6**, 561–568.
- Suryanarayanan, T. S., Murali, T. S., Thirunavukkarasu, N., Govinda Rajulu, M. B., Venkatesan, G. and Sukumar, R., Endophytic fungal communities in woody perennials of three tropical forest types of the Western Ghats, southern India. *Biodiver. Conserv.*, 2011, **20**, 913–928.
- Pandey, A. K., Reddy, M. S. and Suryanarayanan, T. S., ITS-RFLP and ITS sequence analysis of a foliar endophytic *Phyllosticta* from different tropical trees. *Mycol. Res.*, 2003, **107**, 439–444.
- Jeewon, R., Liew, E. C. Y. and Hyde, K. D., Phylogenetic evaluation of species nomenclature of *Pestalotiopsis* in relation to host association. *Fungal Divers.*, 2004, **17**, 39–55.
- Murali, T. S., Suryanarayanan, T. S. and Geeta, R., Endophytic *Phomopsis* species: host range and implications for diversity estimates. *Can. J. Microbiol.*, 2006, **52**, 673–680.
- Wei, J. G., Xu, T., Guo, L.D., Liu A. R., Zhang, Y. and Pan, X. H., Endophytic *Pestalotiopsis* species associated with plants of Podocarpaceae, Theaceae and Taxaceae in southern China. *Fungal Divers.*, 2007, **24**, 55–74.
- Tejesvi, M. V., Tamhankar, S. A., Kini, K. R., Rao, V. S. and Prakash, H. S., Phylogenetic analysis of endophytic *Pestalotiopsis* species from ethnopharmacologically important medicinal trees. *Fungal Divers.*, 2009, **38**, 167–183.
- Govindarajulu, M. B., Thirunavukkarasu, N., Babu, A. G., Aggarwal, A., Suryanarayanan, T. S. and Reddy, M. S., Endophytic Xylariaceae from the forests of Western Ghats, Southern India: distribution and biological activities. *Mycology*, 2013, **4**, 29–37.
- Woolhouse, M. E., Taylor, L. H. and Haydon, D. T., Population biology of multihost pathogens. *Science*, 2001, **292**, 1109–1112.
- Cord-Landwehr, S., Melcher, R. L. J., Kolkenbrock, S. and Moerschbacher, B., A chitin deacetylase from the endophytic fungus *Pestalotiopsis* sp. efficiently inactivates the elicitor activity of chitin oligomers in rice cells. *Sci. Rep.*, 2016, **6**, Article number: 38018.
- Webb, C. O., Ackerly, D. D. and Kembel, S. W., Phylocom: software for the analysis of phylogenetic community structure and trait evolution. *Bioinformatics*, 2008, **24**, 2098–2100.
- Sieber, T. N., Endophytic fungi in forest trees: are they mutualists? *Fungal Biol. Rev.*, 2007, **21**, 75–89.
- Gilbert, G. S. and Webb, C. O., Phylogenetic signal in plant pathogen-host range. *Proc. Natl. Acad. Sci. USA*, 2007, **104**, 4979–4983.
- Saunders, M. and Kohn, L. M., Evidence for alteration of fungal endophyte community assembly by host defense compounds. *New Phytol.*, 2009, **182**, 229–238.
- Larkin, B. G., Hunt, L. S. and Ramsey, P. W., Foliar nutrients shape fungal endophyte communities in Western white pine (*Pinus monticola*) with implications for white-tailed deer herbivory. *Fungal Ecol.*, 2012, **5**, 252–260.
- Van Bael, S., Estrada, C. and Arnold, A. E., Foliar endophyte communities and leaf traits in tropical trees. In *The Fungal Community: Its Organisation and Role in the Ecosystem* (eds Dighton, J. and White, J. F.), CRC Press, Taylor and Francis, Boca Raton, FL, 2017, pp. 79–94.
- May, R. M., How many species are there on earth? *Science*, 1988, **241**, 1441–1449.
- May, R. M., A fondness for fungi. *Nature*, 1991, **352**, 475–476.
- Suryanarayanan, T. S., Govinda Rajulu, M. B., Thirumalai, E., Reddy, M. S. and Money, N. P., Agni's fungi: heat-resistant spores from the Western Ghats, southern India. *Fungal Biol.*, 2011, **115**, 833–838.
- Govinda Rajulu, M. B., Lai, L. B., Murali, T. S., Gopalan, V. and Suryanarayanan, T. S., Several fungi from fire-prone forests of southern India can utilize furaldehydes. *Mycol. Prog.*, 2014, **13**, 1049–1056.
- Shipunov, A., Newcombe, G., Raghavendra, A. and Anderson, C., Hidden diversity of endophytic fungi in an invasive plant. *Am. J. Bot.*, 2008, **95**, 1096–1108.
- Navaud, O., Barbacci, A., Taylor, A., Clarkson, J. P. and Raffaele, S., Shifts in diversification rates and host jump frequencies shaped the diversity of host range among Sclerotiniaceae fungal plant pathogens. *Mol. Ecol.*, 2018, **27**, 1309–1323.
- Redman, R. S., Dunigan, D. D. and Rodriguez, R. J., Fungal symbiosis from mutualism to parasitism: who controls the outcome, host or invader? *New Phytol.*, 2001, **151**, 705–716.
- Krings, M., Taylor, T. N., Hass, H., Kerp, H., Dotzler, N., Hermesen, E. J., Fungal endophytes in a 400-million-yr-old land plant: infection pathways, spatial distribution and host responses. *New Phytol.*, 2007, **174**, 648–657.
- Mejía, L. C. *et al.*, Pervasive effects of a dominant foliar endophytic fungus on host genetic and phenotypic expression in a tropical tree. *Front. Microbiol.*, 2014, **5**, 479.
- Jacott, C. N., Murray, J. D. and Ridout, C. J., Trade-offs in arbuscular mycorrhizal symbiosis: Disease resistance, growth responses and perspectives for crop breeding. *Agronomy*, 2017, **7**, 4–18.
- Gomes, R. R., Glienke, C., Videira, S. I. R., Lombard, L., Groenewald, J. Z. and Crous, P. W., *Diaporthe*: a genus of endophytic, saprobic and plant pathogenic fungi. *Persoonia: Mol. Phyl. Evol. Fungi*, 2013, **31**, 1–41.
- O'Connell, R. J. *et al.*, Lifestyle transitions in plant pathogenic *Colletotrichum* fungi deciphered by genome and transcriptome analyses. *Nat. Genet.*, 2012, **44**, 1060–1065.

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