Endophytes and weed management: a commentary

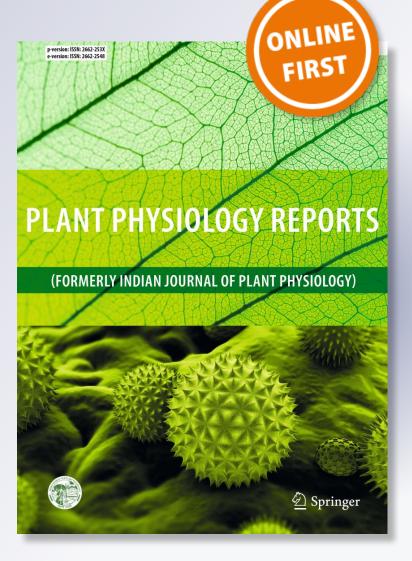
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REVIEW ARTICLE

Endophytes and weed management: a commentary

T. S. Suryanarayanan¹

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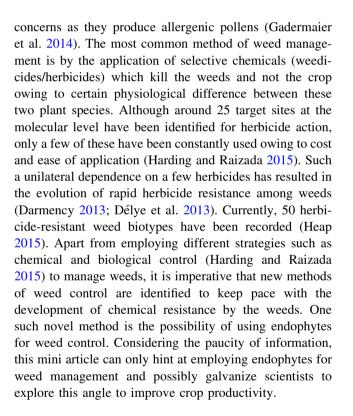
Abstract The inside of plant tissues are the home for some bacteria and fungi. Called the endophytes, these microbes are constantly associated with all plants and constitute their endobiome. Endophytes, probably owing their interaction with their plant hosts, produce many novel biochemicals exhibiting interesting bioactivities. They promote crop growth by increasing nitrogen fixation, hormone production, phosphate and iron utilization. Endophyte association makes the plants more tolerant to pathogens, pests and abiotic stress. As a consequence of such desirable traits, although endophytes have been studied for crop improvement, their possible use in weed management has not been addressed adequately. This mini review cogitates on this facet of endophyte technology.

Keywords Weedicides \cdot Herbicides \cdot Plant microbiome \cdot Endobiome

Introduction

Weeds, under certain conditions cause more economic loss to agriculture than insect pests and pathogenic fungi. Apart from competing with the crop for water, light, nutrients and space, weeds could also harbor pests which attack the crop thereby reducing the yield and increasing production cost. Gharde et al. (2018) estimate that the total economic loss due to weeds in 10 major crops in India is around USD 11 billion. Furthermore, some weeds are responsible for health

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Why endophytes?

Endophytes (bacteria and fungi) which make up the endobiome of plants, have evolved with the plants and together with plant constitute a holobiome. They produce an array of metabolites with novel molecular architecture exhibiting many interesting bioactivities. Endophyte infection of a plant contributes to the plant's resistance (Kang et al. 2007) by the upregulation of hundreds of defense related genes of the plant (Mejía et al. 2014).





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Furthermore, endophytes increase the ecological fitness of their host plants by enhancing their nutrient uptake (Paungfoo-Lonhienne et al. 2010), tolerance to abiotic (Hyde et al. 2019) and biotic (Gond et al. 2015) stressors and increase plant growth and yield (Ryan et al. 2008; Gaiero et al. 2013). These attributes of the endophytes make them an attractive alternative to plant breeding as a method of improving crop traits. Although there are many studies of the species diversity (Kandel et al. 2017; Suryanarayanan et al. 2018a, b) and bioactive metabolites of endophytes (Suryanarayanan et al. 2009), there are only a few investigations addressing the effect of weedicides and herbicides on the endophyte status of plants and the possible production of metabolites by endophytes which may act against weeds. Among the only thirteen authorized bioherbicides currently used, ten are of fungal origin reinforcing the need to screen fungi assiduously for novel weedicides (Samad et al. 2017). With reference to fungi, although their estimated diversity ranges from 2.2 to 3.8 million species, only about 8% of these are currently known (Hawksworth and Lücking 2017); thus exploring fungi, especially the endophytes due to their extraordinary synthetic ability, for novel herbicidal compounds would be worthwhile.

How may endophytes be used for weed management?

A few studies endorse the potential of endophytes in weed control (Kowalski et al. 2015). Considering the high synthetic potential of endophytes (Suryanarayanan et al. 2009), one immediately obvious method of using endophytes to manage weeds is to look for their metabolites which are toxic to weeds. The use of a weedicidal compound secreted by a microbe is a better option in weed control than the use of a biocontrol agent whose efficacy depends on its continued survival in the introduced environment. Schulz et al. (2002) showed that fungal endophytes produce metabolites exhibiting herbicidal activity. Survanarayanan et al. (2018a) reported that fungal endophytes produce metabolites which induce chlorosis followed by necrosis in Lemna minor. Although the authors did not identify the chemicals which induced the death this weed, the study showed that fungal endophytes could be a source of weedicides. Singh et al. (2018) showed that endophytic actinomycetes could be a source of herbicidal metabolites. A Chaetomium globosum isolated as an endophytic fungus from the leaves of Amaranthus viridis produce phytotoxic azaphilone derivatives (Piyasena et al. 2015). Having said that, it is imperative to screen the effective endophyte metabolites for their specificity of action and that they do not act on the crop at the effective concentration. It is also essential to ascertain that the effective chemicals are not mycotoxins as the introduction of such chemicals in the food chain is not desirable. Our study shows that foliar endophytic fungi do produce various mycotoxins (Thirumalai et al. 2013). More detailed investigations are needed addressing interspecific competition among introduced candidate endophytes and the native ones in a plant, and the role and location of each component of the endomicrbiome in plant tissue to fully harness the weedicide potential of endophytes in agriculture (White et al. 2019).

The other side: impacts of herbicides on endophytes

Although endophytes promote crop growth by increasing nitrogen fixation, hormone production, phosphate and iron utilization (Xia et al. 2015), the use of agrochemicals such as pesticides (da Costa Stuart et al. 2018) and fungicides (Mohandoss and Suryanarayanan 2009) alters the endomicrobiome of plants. Treatment of soybean with the herbicide Fenoxaprop-P-ethyl decreased the diversity, richness, and evenness of its fungal endophyte assembly (da Costa Stuart et al. 2018). Similarly, imidazolinone herbicides brought about changes in the composition of fungal endophytes in sugarcane (Stuart et al. 2010). Such alterations in the endobiome could affect negatively the crop performance.

Some bacterial (Rylott 2014) and fungal endophytes (Khan et al. 2014) are known to biotransform many chemicals including xenobiotics and perhaps as a consequence, their species composition in the crop tissue is altered when crops are exposed to chemicals like herbicides. Application of glyphosate herbicide alters the native bacterial endophyte community in soybean by promoting those which could use this chemical as a source of energy and nutrient and eliminating those which are susceptible to it (Kuklinsky-Sobral et al. 2005; Kryuchkova et al. 2014). Wang et al. (2017) showed that Neurospora intermedia, an endophytic fungus isolated from sugarcane roots degrades the phenylurea herbicides diuron, fenuron, monuron, metobromuron, isoproturon, chlorbromuron, and linuron. Many endophytic bacteria and rhizobacteria increase the resistance of their host plants to herbicides by metabolizing them (Liu et al. 2011; Tétard-Jones and Edwards 2016). Inoculation of pea plants with the bacterial endophyte Pseudomonas putida POPHV6 aided in the disappearance of 2,4-D from soil and resulted in reduced translocation of the herbicide in the plant (Germaine et al. 2006). Thus, it is possible that the microbes of the weed endobiome get selected for weedicide tolerance and ultimately add to weed resistance to weedicides. Indeed, Tétard-Jones and Edwards (2016) allude to the possibility of endophytes priming 'the resistance mechanisms in plants such that they enhance herbicide tolerance by inducing the host's stress responses to withstand the downstream toxicity caused by herbicides.' Thus, various aspects evolution of herbicide resistance including the attendant fitness cost which are primarily host gene controlled (Baucom 2019), need to be revisited considering the role played by endophytes.

Although endophytes are known to enhance the abiotic and stress tolerance of plants under laboratory conditions, their role in increasing the fitness of crops may not be as considerable in the field (Serfling et al. 2007). It is known that the differential tolerance to weedicides exhibited by the crop and the weed is due to the difference in the vulnerability of the biochemical process targeted by the weedicide and the relative capacity of these plants to detoxify the weedicide. Recent investigations add a microbial facet to this phenomenon viz. the endobiome of weeds and crops. This warrants focused basic studies on the endophytes, the interaction among them, between their host plant as well as the environment which could ultimately lead to better use of endophytes in weed management.

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References

- Baucom, R. S. (2019). Evolutionary and ecological insights from herbicide-resistant weeds: What have we learned about plant adaptation, and what is left to uncover? *New Phytologist*, 223, 68–82.
- da Costa Stuart, A. K., Stuart, R. M., & Pimentel, I. C. (2018). Effect of agrochemicals on endophytic fungi community associated with crops of organic and conventional soybean (*Glycine max* L. Merril). Agriculture and Natural Resources, 52(4), 388–392.
- Darmency, H. (2013). Pleiotropic effects of herbicide-resistance genes on crop yield: A review. *Pest Management Science*, 69(8), 897–904.
- Délye, C., Jasieniuk, M., & Le Corre, V. (2013). Deciphering the evolution of herbicide resistance in weeds. *Trends in Genetics*, 29(11), 649–658.
- Gadermaier, G., Hauser, M., & Ferreira, F. (2014). Allergens of weed pollen: An overview on recombinant and natural molecules. *Methods*, 66(1), 55–66.
- Gaiero, J. R., McCall, C. A., Thompson, K. A., Day, N. J., Best, A. S., & Dunfield, K. E. (2013). Inside the root microbiome: Bacterial root endophytes and plant growth promotion. *American Journal* of Botany, 100(9), 1738–1750.
- Germaine, K. J., Liu, X., Cabellos, G. G., Hogan, J. P., Ryan, D., & Dowling, D. N. (2006). Bacterial endophyte-enhanced phytoremediation of the organochlorine herbicide 2,4-dichlorophenoxyacetic acid. *FEMS Microbiology Ecology*, 57(2), 302–310.
- Gharde, Y., Singh, P. K., Dubey, R. P., & Gupta, P. K. (2018). Assessment of yield and economic losses in agriculture due to weeds in India. *Crop Protection*, 107, 12–18.

- Gond, S. K., Bergen, M. S., Torres, M. S., White, J. F., & Kharwar, R. N. (2015). Effect of bacterial endophyte on expression of defense genes in Indian popcorn against *Fusarium moniliforme. Symbiosis*, 66(3), 133–140.
- Harding, D. P., & Raizada, M. N. (2015). Controlling weeds with fungi, bacteria and viruses: A review. *Frontiers in Plant Science*, 6, 659.
- Hawksworth, D. L., & Lücking, R. (2017). Fungal diversity revisited: 2.2 to 3.8 million species. *Microbiology Spectrum*, 5(4), 1–2.
- Heap, I. (2015). The International survey of herbicide resistant weeds. Weeds resistant to EPSP synthase inhibitors. Retrieved May 10, 2015.
- Hyde, K. D., Xu, J., Rapior, S., Jeewon, R., Lumyong, S., Niego, A. G. T., et al. (2019). The amazing potential of fungi: 50 ways we can exploit fungi industrially. *Fungal Diversity*, 97(1), 1–136.
- Kandel, S. L., Joubert, P. M., & Doty, S. L. (2017). Bacterial endophyte colonization and distribution within plants. *Microor*ganisms, 5(4), 77.
- Kang, S. H., Cho, H., Cheong, H., Ryu, C. M., Kim, J. F., & Park, S. H. (2007). Two bacterial entophytes eliciting both plant growth promotion and plant defense on pepper (*Capsicum annuum L.*). *Journal of Microbiology and Biotechnology*, 17(1), 96–103.
- Khan, Z., Roman, D., Kintz, T., delas Alas, M., Yap, R., & Doty, S. (2014). Degradation, phytoprotection and phytoremediation of phenanthrene by endophyte *Pseudomonas putida*, PD1. *Envi*ronmental Science and Technology, 48(20), 12221–12228.
- Kowalski, K. P., Bacon, C., Bickford, W., Braun, H., Clay, K., Leduc-Lapierre, M., et al. (2015). Advancing the science of microbial symbiosis to support invasive species management: A case study on *Phragmites* in the Great Lakes. *Frontiers in Microbiology*, 6, 1–14.
- Kryuchkova, Y. V., Burygin, G. L., Gogoleva, N. E., Gogolev, Y. V., Chernyshova, M. P., Makarov, O. E., et al. (2014). Isolation and characterization of a glyphosate-degrading rhizosphere strain, *Enterobacter cloacae* K7. *Microbiological Research*, 169(1), 99–105.
- Kuklinsky-Sobral, J., Araujo, W. L., Mendes, R., Pizzirani-Kleiner, A. A., & Azevedo, J. L. (2005). Isolation and characterization of endophytic bacteria from soybean (*Glycine max*) grown in soil treated with glyphosate herbicide. *Plant and Soil*, 273(1–2), 91–99.
- Liu, Y. J., Liu, S. J., Drake, H. L., & Horn, M. A. (2011). Alphaproteobacteria dominate active 2-methyl-4-chlorophenoxyacetic acid herbicide degraders in agricultural soil and drilosphere. *Environmental Microbiology*, 13(4), 991–1009.
- Mejía, L. C., Herre, E. A., Sparks, J. P., Winter, K., García, M. N., Van Bael, S. A., et al. (2014). Pervasive effects of a dominant foliar endophytic fungus on host genetic and phenotypic expression in a tropical tree. *Frontiers in Microbiology*, 5, 479.
- Mohandoss, J., & Suryanarayanan, T. S. (2009). Effect of fungicide treatment on foliar fungal endophyte diversity in mango. *Sydowia*, 61, 11–24.
- Paungfoo-Lonhienne, C., Rentsch, D., Robatzek, S., Webb, R. I., Sagulenko, E., Näsholm, T., et al. (2010). Turning the table: Plants consume microbes as a source of nutrients. *PLoS ONE*, 5(7), e11915.
- Piyasena, K. G. N. P., Wickramarachchi, W. A. R. T., Kumar, N. S., Jayasinghe, L., & Fujimoto, Y. (2015). Two phytotoxic azaphilone derivatives from *Chaetomium globosum*, a fungal endophyte isolated from *Amaranthus viridis* leaves. *Mycology*, 6(3–4), 158–160.
- Ryan, R. P., Germaine, K., Franks, A., Ryan, D. J., & Dowling, D. N. (2008). Bacterial endophytes: Recent developments and applications. *FEMS Microbiology Letters*, 278(1), 1–9.
- Rylott, E. L. (2014). Endophyte consortia for xenobiotic phytoremediation: The root to success? *Plant and Soil*, 385(1–2), 389–394.

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- Samad, A., Antonielli, L., Sessitsch, A., Compant, S., & Trognitz, F. (2017). Comparative genome analysis of the vineyard weed endophyte *Pseudomonas viridiflava* CDRTc14 showing selective herbicidal activity. *Scientific Reports*, 7(1), 17336.
- Schulz, B., Boyle, C., Draeger, S., Römmert, A. K., & Krohn, K. (2002). Endophytic fungi: A source of novel biologically active secondary metabolites. *Mycological Research*, 106(9), 996–1004.
- Serfling, A., Wirsel, S. G., Lind, V., & Deising, H. B. (2007). Performance of the biocontrol fungus *Piriformospora indica* on wheat under greenhouse and field conditions. *Phytopathology*, 97(4), 523–531.
- Singh, H., Naik, B., Kumar, V., & Bisht, G. S. (2018). Screening of endophytic actinomycetes for their herbicidal activity. *Annals of Agrarian Science*, 16(2), 101–107.
- Stuart, R. M., Romão, A. S., Pizzirani-Kleiner, A. A., Azevedo, J. L., & Araújo, W. L. (2010). Culturable endophytic filamentous fungi from leaves of transgenic imidazolinone-tolerant sugarcane and its non-transgenic isolines. *Archives of Microbiology*, 192(4), 307–313.
- Suryanarayanan, T. S., Thirunavukkarasu, N., Govindarajulu, M. B., Sasse, F., Jansen, R., & Murali, T. S. (2009). Fungal endophytes and bioprospecting. *Fungal Biology Reviews*, 23, 9–19.
- Suryanarayanan, T. S., Devarajan, P. T., Girivasan, K. P., Govindarajulu, M. B., Kumaresan, V., Murali, T. S., et al. (2018a). The host range of multi-host endophytic fungi. *Current Science*, 115(10), 1963–1969.

- Suryanarayanan, T. S., Govinda Rajulu, M. B., & Vidal, S. (2018b). Biological control through fungal endophytes: Gaps in knowledge hindering success. *Current Biotechnology*, 7(3), 185–198.
- Tétard-Jones, C., & Edwards, R. (2016). Potential roles for microbial endophytes in herbicide tolerance in plants. *Pest Management Science*, *72*(2), 203–209.
- Thirumalai, E., Dastjerdi, R., Döll, K., Venkatachalam, A., Karlovsky, P., & Suryanarayanan, T. S. (2013). Mycotoxins of endophytic *Fusarium mangiferae* and *F. pallidoroseum* from betel leaves (*Piper betle* L.). In 35th Mycotoxin workshop, Ghent, Belgium.
- Wang, Y., Li, H., Feng, G., Du, L., & Zeng, D. (2017). Biodegradation of diuron by an endophytic fungus *Neurospora intermedia* DP8-1 isolated from sugarcane and its potential for remediating diuron-contaminated soils. *PLoS ONE*, 12(8), e0182556.
- White, J. F., Kingsley, K. L., Zhang, Q., Verma, R., Obi, N., Dvinskikh, S., et al. (2019). Review: Endophytic microbes and their potential applications in crop management. *Pest Management Science*, 75, 2558–2565.
- Xia, Y., DeBolt, S., Dreyer, J., Scott, D., & Williams, M. A. (2015). Characterization of culturable bacterial endophytes and their capacity to promote plant growth from plants grown using organic or conventional practices. *Frontiers in Plant Science*, 6, 490.

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